Investigation of Possible Positive Effects of Arbutin Application in Experimental Colitis Model

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ABSTRACT

Background/Aims: This study aimed to investigate the possible positive effects of arbutin in a trinitrobenzene sulfonic acid (TNBS)induced experimental colitis model, to compare it with mesalazine, which is used in treating inflammatory bowel disease and to observe the effect of its concomitant use.

Materials and Methods: Forty Wistar albino species male rats were randomized into 5 groups as control, colitis, colitis + arbutin (Arb), colitis + mesalazine (Mes), and colitis + mesalazine + arbutin (M+A). Proinflammatory cytokines [interleukin (IL)-6, IL-1 β , tumor necrosis factor alpha (TNF- α)] and oxidant/antioxidant parameters [malondialdehyde (MDA), superoxide dismutase inhibition (SOD) inhibition, myeloperoxidase (MPO), and catalase, glutathione peroxidase (GPx)] were processed from the samples. Histopathological evaluation evaluated goblet cell reduction, cellular infiltration, and mucosal loss.

Results: When the treatment groups and the TNBS group were compared, statistical significance was achieved in MDA, MPO, SOD inhibition, GPx values, IL-6, IL-1β and TNF-α levels. Histopathological evaluation revealed a statistically significant decrease in the mucosal loss value in the group where mesalazine and arbutin were used together compared to the TNBS group.

Conclusion: Our study's results elaborated that using arbutin alone or in combination with mesalazine produced positive effects in colitis-induced rats.

Keywords: Arbutin, experimental animal model, mesalazine, ulcerative colitis

INTRODUCTION

Ulcerative colitis (UC) is an inflammatory disease that affects the gastrointestinal tract becomes chronic and progresses with relapses. Its etiology and pathogenesis still need to be fully elucidated. However, in recent years, significant progress has been made in revealing the pathogenesis of the disease. Genetics, immunological factors, intestinal flora, and environmental characteristics are thought to play a role in the etiology. Treatment of UC is non-specific. In general, it is based on agents with antiinflammatory activity.^{1,2} The main goals of treatment in UC are to achieve remission, increase the patient's quality of life, and prevent complications that may develop.³ Experimental colitis models, particularly 2,4,6 trinitrobenzene sulfonic acid (TNBS), have been utilized in various experimental animal models; a single fast, reliable, and reproducible method is essential for exhibiting pharmacological approaches to UC.4

Experimental and clinical studies show that triggering oxidative stress advances the development of IBD. There is a correlation between the increase of reactive oxygen species in UC and the degree of inflammation. Oxidative stress damages the gastrointestinal mucosa and bacterial invasion, stimulating the immune response and initiating IBD.^{5,6}

Anti-inflammatory agents and antioxidant defense mechanisms positively affect the course of IBD by preventing the initiation of lipid peroxidation, forming free radicals, and increasing cytokines.⁷ Mesalazine, 5-aminosalicylic acid (5-ASA), is the standard for treating uncomplicated, mild to moderate UC. Meta-analyses of randomized controlled trials have shown the superiority of mesalazine over placebo and rectal steroids in both remission and maintenance therapy in the treatment of UC.

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Mesalazine can be administered rectally through suppositories, foams, or enemas.^{8,9} It is a β -glucoside derived from arbutin hydroguinone (HQ; 1,4-dihydroxybenzene), which is obtained from the leaves of various plants, such as blueberries. The leaves of these plants have traditionally been used for many years by the natives of China and the American continent as folk remedies, especially for wound healing and treating urinary tract infections.¹⁰⁻¹⁵ Arbutin has been shown to have long-term free radical removal properties and protective properties from oxidative stress by inhibiting lipid peroxidation.^{10,13,16} Studies on arbutin have shown that it increases the antiinflammatory effects of corticosteroids and indomethacin. In addition, arbutin reduces free radical formation in neutrophils and decreases enzyme release from neutrophils in experimental arthritis and in vitro studies.17 Experimental studies on intestinal epithelial cells have shown that arbutin increases cell viability.¹⁸

Although oxidative stress and antioxidant power level are in balance under physiological conditions, this balance is disrupted in the direction of oxidative damage in UC processes, disrupting the occurring inflammation process, increasing tissue damage, and prolonging the healing period. Therefore, biochemical analysis of oxidative damage and antioxidant biomarkers glutathione peroxidase (GPx), malondialdehyde (MDA), superoxide dismutase inhibition (SOD), myeloperoxidase (MPO), and catalase (CAT) levels, together with histopathological evaluation, reveals the positive efficacy of treatments applied against ulcerative colitis at the tissue level.

The study aimed to compare the positive effect of arbutin against induced UC with routine mesalazine therapy and combined therapy.

MATERIALS AND METHODS

This study was carried out in Aydın Adnan Menderes University Faculty of Medicine Experimental Animals Laboratory using a total of 40 Wistar albino species male rats with weights ranging from 250-400 g obtained from Aydın Adnan Menderes University Faculty of Medicine Experimental Animals Laboratory (ethical approval:

Main Points

- Arbutin has anti-inflammatory and antioxidant effects.
- Arbutin has a protective and therapeutic effect on colitis.
- Colon structure deteriorated with trinitrobenzene sulfonic acid is healed by arbutin administration.

HADYEK dated May 27, 2021, and 64583101/2021/073), biochemical analyses were performed in Aydın Adnan Menderes University Faculty of Medicine Physiology Laboratory, and histopathological analyzes were performed in Aydın Adnan Menderes University Faculty of Medicine Pathology Laboratory. The animals were allowed to be in pre-experimental cages in a controlled room with a 12-hour light–darkness cycle. They were fed with standard rat feed and allowed to drink water from the fad. After the environment was adapted, all the animals were starved 12 hours before the experiment, and the experiment was started (Table 1).

Sham Control Group

Rats in this group were not exposed to colitis. Saline was administered intragastric and rectal by cannula.

Colitis Group

The intestines of rats in this group and treatment groups were emptied, under ketamine (90 mg/kg) and xylazine (10 mg/kg) anesthesia, diluted in 0.8 mL saline of 25 mg of TNBS. The solution, prepared by dissolving in 37% ethanol, was given by entering 8 cm inside the anal orifice with the help of a polyethylene cannula.^{19,20} No therapeutic agents were administered to rats in the colitis group.

Colitis + Mesalazine Group

The first 100 mg/kg mesalazine administration was performed intragastric 3 times, 6 hours after colitis first gavage was performed.

Colitis + Arbutin Group

Arbutin 250 mg/kg was administered intragastrically 3 times, the first of which was 6 hours after colitis first gavage was performed.

Colitis + Mesalazine + Arbutin Group

In the first one, 6 hours after the formation of colitis, 3 times 100 mg/kg mesalazine and 250 mg/kg arbutin administration were performed intragastric route.

Colitis was formed in all rats in the treatment groups, and no therapeutic agents were given before colitis was formed. Induction of experimental colitis was performed by applying the procedure in the colitis group. Seventytwo hours after colitis was formed, rats were sacrificed under ketamine xylazine anesthesia. At the end of the process, colon tissue was taken from the rats, and the experiment was terminated.

	Hour 0	Hour 6	Hour 30	Hour 54	Hour 72
Sham control	Intrarectal saline	First saline gavage	Second saline gavage	Third saline gavage	Sacrification
TNBS	Colitis induced	-	-	-	Sacrification
Mes	Colitis induced	First Mes gavage	Second Mes gavage	Third Mes gavage	Sacrification
Arb	Colitis induced	First Arb gavage	Second Arb gavage	Third Arb gavage	Sacrification
Mes+arb	Colitis induced	First Mes + arb gavage	Second Mes + arb gavage	Third Mes + arb gavage	Sacrification
Arb, arbutin; Mes, mesalazine; TNBS, trinitrobenzene sulfonic acid.					

 Table 1. Experimental Diagram

The tissues for biochemical analysis were quickly taken to a -80° C freezer and preserved until the analysis day. On the day of the analysis, homogenization of the tissues in phosphate buffer (1 : 20) was performed, and procedures were performed in accordance with the instructions inside the kits. Kits (BioVision, Milpitas, Calif, USA) was used to measure tissue GPx, MDA, SOD, MPO, and CAT levels. Tissue samples were followed by instructions with ScienceCell (Calif, USA) brand rat ELISA kits to measure interleukin-1 beta (IL-1 β), interleukin-6 (IL-6), and tumor necrosis factor alpha (TNF- α) levels. All tests were analyzed with the help of the "Diagnostic Automation, Inc./ ELx800TM" brand microplate reader.

The tissues were detected in 10% neutral buffered formalin and taken to routine tissue follow-up. After this procedure, 4 μ m thick sections were prepared with rotary microtomes from tissue samples embedded in paraffin blocks. These sections were stained with hematoxylin eosin (HE) and evaluated at magnifications of ×4, ×10, ×20, and ×40 under the light microscope (BX51, Olympus, Tokyo, Japan). The evaluation was made by a single pathologist blindly without knowing the groups in the study. In the evaluation, semiquantitative scoring was performed according to the criteria used in an earlier study by Appleyard and Wallace (Table 2).²¹

Statistical Analysis

All statistical evaluation of our study was performed using GraphPad Prism (version 7.04, GraphPad Software

Inc., San Diego, Calif, USA) program. One-way analysis of variance was used for statistical evaluation between groups. In the interpretation of the values and the multiple comparisons of the group values, "Tukey's multiple comparisons," "Dunnett's multiple-comparisons," and "Sidak's multiple comparisons" tests were applied. The values of the groups were expressed as standard deviations \pm mean, and P < .05 was considered statistically significant. Excel (Microsoft, Wash, USA) program was used to prepare the graph showing the weight change and to determine the average values. The distributions of biochemical measurements were evaluated by the Shapiro–Wilk's test and skewness/kurtosis statistics.

RESULTS

Catalase level results showed the lowest values in the colitis group, although there was no statistically significant difference between the control group and the colitis and treatment groups (P > .05) (Figure 1A).

Increase has been observed in colon GPx levels in all treatment groups compared to the TNBS group (P < .05). We found an increase in GPx level (P < .005) compared to the TNBS group in the group that received mesalazine alone, a significant increase in GPx level compared to the TNBS group in the group that underwent mesalazine + arbutin (P < .01), and increase in the GPx level compared to the TNBS group in the group that received arbutin-alone (P < .05) (Figure 1B).

Table 2. Scoring Criteria Used in the Pathological Microscopic Evaluation

Score	Loss of Mucous Membranes	Cellular Infiltration	Crypt Abscess	Goblet Cell Reduction (Necrosis/Ulceration)
0	No	No	No	No
1	Under 5%	Few	Apparent	Apparent
2	5%-10%	Middle	-	-
3	Over 10%	Apparent	-	-



Figure 1. (A) Distribution of catalase values in study groups, (B) distribution of glutathione peroxidase values in study groups, (C) distribution of superoxide dismutase inhibition inhibition values in study groups, and (D) distribution of myeloperoxidase values in study groups. Arb, arbutin; Mes, mesalazine; TNBS, trinitrobenzene sulfonic acid.

We found a statistically significant decrease in SOD inhibition in all treatment groups compared to the TNBS group (P < .05). This difference was significant in the group applied together with arbutin+mesalazine (P < .005) (Figure 1C).

Colon MPO levels were found to be highest in the TNBS group (P < .001). Both treatments applied caused a significant reduction, but there was no statistical difference between them (P > .05) (Figure 1D).

We found a decrease in MDA in all treatment groups compared to the TNBS group (P < .005). This significance



Figure 2. (A) Distribution of malondialdehyde values in working groups, (B) distribution of interleukin (IL)-1 β values in working groups, (C) distribution of IL-6 values in working groups, (D) distribution of tumor necrosis factor alpha values in study groups. Arb, arbutin; Mes, mesalazine; TNBS, trinitrobenzene sulfonic acid.

is highest in arbutin + mesalazine group (P < .001) (Figure 2A).

We found a statistically significant decrease in tissue IL-1 β levels in treatment groups against TNBS group (P < .01). When the treatment groups were compared among themselves, there was no significant difference in IL-1 β levels between the treatment groups (P > .05) (Figure 2B).

There was a statistically significant decrease in tissue IL-6 levels in all treatment groups compared to the TNBS group (P < .005). This difference was more significant in the group that received mesalazine alone (P < .001) (Figure 2C).

	Control	TNBS	Mes	Arb	M+A
MPO (mU/mL)	373000 ± 71621	701000 ± 61031	410333 ± 39807	385833 ± 66406	357167 ± 81263
CAT (mU/mL)	126167 ± 27088	102833 ± 17394	93666 ± 19551	96666 ± 20412	99000 ± 21410
GPx (mU/mL)	870667 ± 83377	412167 ± 38602	605000 ± 81545	530167 ± 75358	570667 ± 55164
IL-1β (pg/g)	1092670 ± 129728	1613670 ± 163867	1152170 ± 226414	1102500 ± 256475	1109000 ± 186654
IL-6 (pg/g)	6950 ± 1059	13083 ± 2235	7866 ± 1032	8500 ± 1663	8383 ± 1479
MDA (nmol/g)	71833 ± 19446	240333 ± 39195	146167 ± 35391	153833 ± 30393	127000 ± 24025
SOD inhibition (%)	21166 ± 7082	56000 ± 12280	39833 ± 5845	40333 ± 7501	31000 ± 7375
TNF-α (pg/g)	1179330 ± 140773	2014650 ± 242701	1386670 ± 157504	1425920 ± 219728	1245830 ± 148817

Arb, arbutin; CAT, catalase; GPx, glutathione peroxidase; IL, interleukin; MDA, malondialdehyde; Mes, mesalazine; MPO, myeloperoxidase; SOD, superoxide dismutase inhibition; TNBS, trinitrobenzene sulfonic acid; TNF-α, tumor necrosis factor-alpha.

Table 3. Biochemical Parameter Values

The decrease in tissue TNF- α was statistically significant in all treatment groups compared to the TNBS group (P < .005). This difference was significant in the groups treated together with mesalazine and arbutin+mesalazine alone (P < .001) (Figure 2D).

All biochemical results were given in Table 3.

In histopathological tissue analysis, no statistically significant decrease in goblet cell reduction in the treatment groups against the TNBS group (P > .05) has been detected. Although there was an observational difference in the group in which mesalazine and arbutin were used together, there was no statistically significant difference (P > .05) (Figure 3A). Although cellular infiltration values were observed in histopathological evaluation compared to the TNBS group in the treatment groups, there was no significance (P > .05) (Figure 3B). In the histopathological evaluation of mucosal losses, although there was an observational difference in the arbutin and mesalazine groups alone compared to the TNBS group, there was no statistically significant decrease (P > .05). In the group where mesalazine and arbutin were used together, we found



Figure 3. (A) Distribution of goblet cell reduction values in study groups. (B) Distribution of cell infiltration values in study groups. (C) Distribution of mucosal loss values in study groups.

a statistically significant decrease in the mucosal loss value compared to the TNBS group (P < .05) (Figure 3C). Histopathological microscopic images are given in Figure 4.



Figure 4. (A) Column with a normal appearance in the control group (HE, ×40). (B) Colon with extensive ulceration (above 10%), lyricin cellular infiltration, and Goblet cell reduction in the TNBS group (HE, ×40). (C) Ulceration (5%-10%) in the mesalazine-treated group, colon with moderate cellular infiltration (HE, ×40). (D) Ulceration (5%-10%) in the Arbutin-treated group, colon with moderate cellular infiltration (HE, ×40). (E) Ulceration (less than 5%) in the group treated with mesalazine and arbutin, colon with minimal cellular infiltration (HE, ×40).

Table 4. Histological Parameter Values						
	Control	TNBS	Mes	Arb	M+A	
Goblet cell reduction	0.000 ± 0.000	1.000 ± 0.000	1.000 ± 0.000	1.000 ± 0.000	0.625 ± 0.517	
Cellular infiltration	0.375 ± 0.517	2.500 ± 0.534	2.375 ± 0.517	2.125 ± 0.353	$2.000\pm0,000$	
Mucosal loss	0.000 ± 0.000	2.875 ± 0.353	2.375 ± 0.517	2.375 ± 0.517	1.500 ± 0.534	
Arb, arbutin; Mes, mesalazi	ne; TNBS, trinitrobenzene	e sulfonic acid.				

All histopathological results are given in Table 4.

DISCUSSION

In our study, we investigated the possible positive effects of arbutin in the TNBS-induced experimental colitis model. We compared it with mesalazine used in the treatment of IBD and observed the effect of using it together. Proinflammatory cytokines (IL-6, IL-1β, TNF- α) and oxidant/antioxidant parameters (MDA, MPO, SOD inhibition, CAT, GPx) were processed in the study groups. In histopathological evaluation, goblet cell reduction, cellular infiltration and mucosal loss were evaluated. When the treatment groups were compared with the TNBS group, statistical significance was achieved in MDA, MPO, SOD inhibition, GPx values, IL-6, IL-1 β , and TNF- α levels. In histopathological evaluation, a statistically significant decrease in mucosal loss was observed in the group where mesalazine and arbutin were used together compared to the TNBS group.

In our study, mesalazine, arbutin and mesalazine + arbutin were given 6 hours after TNBS application. Pontell et al²² in their study, showed that inflammation began 3 hours after TNBS injection into the ileal lumen and that the immediate damage caused by TNBS in the mucosa was rapidly reversed (<1 day).

In our we study found a statistically significant decrease in tissue MDA value in all treatment groups compared to the TNBS group. In the in vitro study of Takebayashi et al¹⁶ with arbutin, they reported that arbutin exerts antioxidant activity comparable to, or even more potent, than hydroquinone. In the gentamicin-induced experimental nephrotoxicity study conducted by Emadi et al,²³ it was reported that the administration of arbutin prevented gentamicin-induced nephrotoxicity, and microscopic, oxidant, and antioxidant parameters supported the results. In our study, arbutin's antioxidant efficacy was determined by biochemical and histopathological analyses. In the Alzheimer's model conducted by Dastan et al,²⁴ MDA levels decreased in the group that used arbutin as a preservative. In our study, although tissue CAT levels were higher in the control group compared to colitis and treatment groups, there was no statistically significant difference. In a study conducted by Seyfizadeh et al²⁵ with arbutin in Hep-G2 cells, it was observed that the administration of arbutin increased CAT and SOD levels and decreased MDA levels. In an experimental ethanol-induced liver injury model study conducted by Wang et al,²⁶ a significant increase in SOD and GPx levels was found in rats treated with arbutin, and it was also shown that the levels of the inflammatory cytokines TNF- α and IL-6 decreased significantly in rats treated with arbutin.

We found a statistically significant decrease in MDA value in all treatment groups compared to the TNBS group. As for the SOD inhibition value, there was a statistically significant decrease in all treatment groups compared to the TNBS group, consistent with the increase in SOD levels shown in the studies.

Our study found a statistically significant increase in tissue GPx levels in all treatment groups compared to the TNBS group and a statistically significant decrease in TNF- α , IL-6, and IL-1 β levels in all treatment groups compared to the TNBS group. In the lipopolysacchar ide-induced experimental lung injury model conducted by Ye et al²⁷ in rats, it was observed that the level of SOD increased and the MDA level decreased in the rats treated with arbutin compared to the control group. The inflammatory cytokines TNF- α , IL-6, and IL-1 β were reduced in arbutin-treated mice compared to the control group.

In Bian et al's²⁸ experimental model of lipopolysaccharide-induced sepsis pneumonia in mice, it was observed that MPO and MDA levels decreased in mice treated with arbutin and levels of the inflammatory cytokines TNF- α , IL-6, and IL-1 β in mice treated with arbutin. These results support our study and show that oxidative damage markers and inflammatory parameters are reduced.

In our study, no therapeutic agent was given before the formation of experimental colitis, and arbutin administration

of 250 mg/kg was performed intragastric 3 times, the first of which was 6 hours after the experimental colitis was formed. Although histopathological examination revealed observational differences in goblet cell reduction and cellular infiltration in the treatment groups compared to the TNBS group, there was no statistically significant difference. A statistically significant decrease in the mucosal loss value was found in the group using mesalazine and arbutin together compared to the TNBS group. In the experimental study in which the gastroprotective property of arbutin was examined by Taha et al,²⁹ 30 and 60 mg/kg arbutin was administered orally for 14 days. This study observed decreased ulcer area, submucosal edema, and leukocyte infiltration in rats treated with arbutin. The MDA levels in the stomach tissue decreased in rats treated with arbutin. The study observed that TNF and IL-6 levels increased in ulcerated rats compared to the control group, while they did not increase in rats treated with arbutin.

Our study found that TNF- α , IL-6, and IL-1 β levels decreased statistically significantly in all treatment groups compared to the TNBS group. These results show that the anti-inflammatory property of arbutin is effective on the intestinal mucosa. In our study, it is essential that the anti-inflammatory properties of arbutin as well as the antioxidant properties of arbutin, were shown in the intestinal tissue and that arbutin and mesalazine were administered together and compared with other groups. Unlike other studies, the fact that we have administered arbutin in 3 repeated doses instead of in the form of low doses for a long time in the dose of 250 mg/kg may also be a guide for the use in the acute treatment of IBD exacerbations. In a dextran sodium sulfate (DSS)-induced experimental colitis model study conducted by Zhang et al³⁰ in mice, intragastric arbutin was administered in doses of 50 mg/kg and 100 mg/kg for 7 days to mice with experimental colitis. In histopathological examination, The total histological colitis score was calculated by scoring the severity, degree of inflammation, crypt damage, and participation in percentage. Histopathological improvement was detected in the groups treated with arbutin. Wang et al³¹ conducted a DSS-induced experimental colitis model study in mice that evaluated the anti-inflammatory properties of arbutin. In the study, intragastric arbutin was administered to mice with experimental colitis for 7 days at doses of 50 mg/kg and 100 mg/kg. The results were evaluated by comparing them with sulfasalazine. Histopathological examination evaluated goblet cell loss, crypt loss, epithelial damage, and mucosal infiltration and scored. In the group treated with

arbutin, histopathological examination showed significant improvement. In our study, unlike these 2 studies, we created the experimental colitis model with TNBS. We used mesalazine instead of the sulfasalazine Wang et al³¹ used and compared it to arbutin. We also compared it with other groups by applying arbutin and mesalazine together. The total dose of arbutin administered in our study was similar to the maximum total dose of Wang et al.³¹ Although histopathological examination revealed observational differences in goblet cell reduction and cellular infiltration in the treatment groups compared to the TNBS group, there was no statistically significant difference. We found a statistically significant decrease in the mucosal loss value in the group using mesalazine and arbutin together compared to the TNBS group. The study by Wang et al³¹ calculated and evaluated the total histological score. Our study evaluated goblet cell reduction, cellular infiltration, and mucosal loss values separately. The difference in the results of the histopathological examination depends on this.

Although the model used in the study is important for the healing processes of the colon since it is based on TNBS damage, it does not carry all the pathophysiological details of the UC process. In our study, one of the limitations of the study is that sacrification was not performed at the sixth hour to confirm whether colitis occurred within 6 hours as a result of TNBS induction in rats and that the number of groups and animals in the groups was limited.

Although the use of rats of both genders in our experimental modeling would be of great importance in revealing the healing process and effect differences between the genders, it was determined that male rats/mice were especially preferred in experimental studies to be hormonally stable and to eliminate the effects of hormonal changes on the healing processes.^{28,29,31} Male rats were used in the study because the parameters examined in our study focused on the intergroup differences between the treated and untreated rats and the untreated rats regardless of gender. Only male rats were used in the study, and therefore, the inability to make comparisons between genders and the inability to molecularly examine the biochemical parameters examined at the transcription and translation level due to limited budget are limitations.

In conclusion, in our study, it was thought that arbutin had a protective antioxidant effect on SOD, MDA, and MPO in colitis-induced rats. Statistically significant decrease in TNF- α , IL-6, and IL-1 β values in groups treated with arbutin shows the anti-inflammatory efficacy of arbutin. In our study, biochemical and histopathological results showed that arbutin administration revealed positive results in colitis-induced rats. The use of arbutin alone or in combination with mesalazine in UC is promising for the future in terms of treatment. We think further studies on this subject will help understand the therapeutic properties of arbutin on UC.

Ethics Committee Approval: This study was approved by Ethics Committee of Aydin Adnan Menderes University (approval no: 64583101/2021/073; date: May 27, 2021).

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REFERENCES

1. Guan Q. A comprehensive review and update on the pathogenesis of inflammatory bowel disease. J Immunol Res. 2019;2019:7247238. [CrossRef]

2. Monteleone G, Fina D, Caruso R, Pallone F. New mediators of immunity and inflammation in inflammatory bowel disease. Curr Opin Gastroenterol. 2006;22(4):361-364. [CrossRef]

3. Kornbluth A, Sachar DB. Gastroenterology PPCotACo. Ulcerative Colitis Practice Guidelines in Adults: American College of Gastroenterology, Practice Parameters Committee. 2010;105(3):501-523.

4. Silva I, Pinto R, Mateus V. Preclinical study in vivo for new pharmacological approaches in inflammatory bowel disease: a systematic review of chronic model of TNBS-induced colitis. J Clin Med. 2019;8(10):1574. [CrossRef]

5. Goyette P, Labbé C, Trinh TT, Xavier RJ, Rioux JD. Molecular pathogenesis of inflammatory bowel disease: genotypes, phenotypes and personalized medicine. Ann Med. 2007;39(3):177-199. [CrossRef]

6. Sedghi S, Fields JZ, Klamut M, et al. Increased production of luminol enhanced chemiluminescence by the inflamed colonic mucosa in patients with ulcerative colitis. Gut. 1993;34(9):1191-1197. [CrossRef]

7. Williams JG, Hughes LE, Hallett MB. Toxic oxygen metabolite production by circulating phagocytic cells in inflammatory bowel disease. Gut. 1990;31(2):187-193. [CrossRef]

8. Wang Y, Parker CE, Bhanji T, Feagan BG, MacDonald JK. Oral 5-aminosalicylic acid for induction of remission in ulcerative colitis. CDSR. Cochrane Database Syst Rev. 2016;2016(4):CD000543. [CrossRef]

9. Feagan BG, MacDonald JK. Oral 5-aminosalicylic acid for maintenance of remission in ulcerative colitis. CDSR. Cochrane Database Syst Rev. 2012;10:CD000544. [CrossRef]

10. Carmen P, Vlase L, Tamas M. Natural resources containing arbutin. Determination of arbutin in the leaves of Bergenia crassifolia (L.) Fritsch. acclimated in Romania. Not Bot Horti Agrobot. 2009;37(1): 129-132.

11. Lindpaintner E. Arbutin und Methylarbutin und ihre Bestimmung in Drogen. Mit Unterstützung der Deutschen Forschungsgemeinschaft. Arc Pharm. 1939;277(9):398-415. [CrossRef]

12. Yamaha T, Cardini CE. The biosynthesis of plant glycosides. I. Monoglucosides. Arch Biochem Biophys. 1960;86(1):127-132. [CrossRef]

13. Garrett JT. The Cherokee Herbal: Native Plant Medicine from the Four Directions. Simon and Schuster; 2003.

14. Garcia-Jimenez A, Teruel-Puche JA, Berna J, Rodriguez-Lopez JN, Tudela J, Garcia-Canovas F. Action of tyrosinase on alpha and beta-arbutin: a kinetic study. PLoS One. 2017;12(5):e0177330. [CrossRef]

15. Xu WH, Liang Q, Zhang YJ, Zhao P. Naturally occurring arbutin derivatives and their bioactivities. Chem Biodivers. 2015;12(1):54-81. [CrossRef]

16. Takebayashi J, Ishii R, Chen J, Matsumoto T, Ishimi Y, Tai A. Reassessment of antioxidant activity of arbutin: multifaceted evaluation using five antioxidant assay systems. Free Radic Res. 2010;44(4):473-478. [CrossRef]

17. Pečivová J, Nosál' R, Sviteková K, Mačičková T. Arbutin and decrease of potentially toxic substances generated in human blood neutrophils. Interdiscip Toxicol. 2014;7(4):195-200. [CrossRef]

18. Zhou W, Chen K, Lu Q, et al. The protective effect of rosavin from Rhodiola rosea on radiation-induced intestinal injury. Chem Biodivers. 2020;17(12):e2000652. [CrossRef]

19. Ek RO, Serter M, Ergin K, et al. The effects of caffeic acid phenethyl ester (CAPE) on TNBS-induced colitis in ovariectomized rats. Dig Dis Sci. 2008;53(6):1609-1617. [CrossRef]

20. Deniz M, Çetinel S, Kurtel H. Blood flow alterations in TNBSinduced colitis: role of endothelin receptors. Inflamm Res. 2004;53(7):329-336. [CrossRef]

21. Appleyard CB, Wallace JL. Reactivation of hapten-induced colitis and its prevention by anti- inflammatory drugs. Am J Physiol. 1995;269(1):G119-G125. [CrossRef]

22. Pontell L, Castelucci P, Bagyánszki M, et al. Structural changes in the epithelium of the small intestine and immune cell infiltration of enteric ganglia following acute mucosal damage and local inflammation. Virchows Arch. 2009;455(1):55-65. [CrossRef]

23. Emadi E, Pouramir M, Ghasemi-Kasman M, Feizi F, Halalkhor S, Moghadamnia AA. Arbutin attenuates nephrotoxicity induced by gentamicin. Avicenna J Phytomed. 2021;11(3):210-217.

24. Dastan Z, Pouramir M, Ghasemi-Kasman M, et al. Arbutin reduces cognitive deficit and oxidative stress in animal model of Alzheimer's disease. Int J Neurosci. 2019;129(11):1145-1153. [CrossRef]

25. Seyfizadeh N, Tazehkand MQ, Palideh A, et al. Is arbutin an effective antioxidant for the discount of oxidative and nitrosative stress in Hep-G2 cells exposed to tert-butyl hydroperoxide? Bratisl Lek Listy. 2019;120(8):569-575. [CrossRef]

26. Wang R, Mu J. Arbutin attenuates ethanol-induced acute hepatic injury by the modulation of oxidative stress and Nrf-2/HO-1signaling pathway. JBiochem Mol Toxicol. 2021;35(10):e22872. [CrossRef]

27. Ye J, Guan M, Lu Y, Zhang D, Li C, Zhou C. Arbutin attenuates LPS-induced lung injury via Sirt1/Nrf2/NF-κBp65 pathway. Pulm Pharmacol Ther. 2019;54:53-59. [CrossRef]

28. Bian XX, Zhao X, Ma CH, Shen CP. Arbutin alleviates LPS induced sepsis pneumonia in mice. Evid Based Complement Alternat Med. 2022;2022:5863952. [CrossRef]

29. Taha MME, Salga MS, Ali HM, Abdulla MA, Abdelwahab SI, Hadi AHA. Gastroprotective activities of Turnera diffusa Willd. ex Schult. revisited: role of arbutin. J Ethnopharmacol. 2012;141(1):273-281. [CrossRef]

30. Zhang C, Zhu H, Jie H, Ding H, Sun H. Arbutin ameliorated ulcerative colitis of mice induced by dextran sodium sulfate (DSS). Bioengineered. 2021;12(2):11707-11715. [CrossRef]

31. Wang L, Feng Y, Wang J, et al. Arbutin ameliorates murine colitis by inhibiting JAK2 signaling pathway. Front Pharmacol. 2021;12: 683818. [CrossRef]